

Crystallization and Preliminary X-Ray Diffraction Studies of *N*-Acyl-D-glucosamine 2-Epimerase from Porcine Kidney

Isafumi Maru,* Jun Ohnishi,* Yasuhiro Ohta,* Wataru Hashimoto,[†] Yoji Tsukada,* Kousaku Murata,^{†,1} and Bunzo Mikami[†]

*Kyoto Research Laboratories, Marukin Shoyu Co., Ltd., Uji, Kyoto 611; and [†]Research Institute for Food Science, Kyoto University, Uji, Kyoto 611

Received for publication, July 11, 1996

N-Acyl-D-glucosamine 2-epimerase from porcine kidney, which was cloned and expressed in *Escherichia coli*, was crystallized by the vapor-diffusion method, using polyethylene glycol and ammonium acetate as precipitants. The crystals were resistant to X-ray radiation damage and diffracted to more than 2.0 Å resolution. The diffraction pattern indicated that the crystals belong to the orthorhombic system, space group *P*2₁2₁2₁, with unit-cell dimensions of *a*=78.1, *b*=97.2, and *c*=100.7 Å. It is supposed that the asymmetric unit consists of two *N*-acyl-D-glucosamine 2-epimerase molecules. Collection of data on the native crystals indicated that they are suitable for X-ray structural analysis.

Key words: *N*-acyl-D-glucosamine 2-epimerase, crystallization, porcine kidney, renin-binding protein, X-ray crystallography.

An *N*-acyl-D-glucosamine 2-epimerase (GlcNAc 2-epimerase) [EC. 5.1.3.8] that catalyzes the conversion of *N*-acetyl-D-glucosamine (GlcNAc) to *N*-acetyl-D-mannosamine (ManNAc) has been found in porcine kidney, and rat kidney, liver, spleen, brain, intestinal mucosa, thymus, pancreas, and salivary gland (1, 2). Ghosh and Roseman (2), and Datta (3) partially purified the enzyme from porcine kidney and found that its activity is modulated by a catalytic amount of ATP. ATP was not essential for the enzyme reaction, but the activity of the enzyme was enhanced about 20-fold in the presence of ATP or deoxy-ATP (2, 4). Datta (3) also reported that GlcNAc 2-epimerase possesses two distinct interaction sites, a catalytic site for the substrate and an allosteric site for ATP. We have purified GlcNAc 2-epimerase to homogeneity from porcine kidneys and cloned the gene for the enzyme in *Escherichia coli* (4). The enzyme is a dimer of identical subunits, each with a molecular weight of about 46 kDa. The nucleotide sequence of the cDNA was determined, and the predicted 402 amino acid residues were characterized by the presence of a leucine zipper motif and by the absence of a hydrophobic amino-terminal sequence indicative of a signal sequence. There is a potential asparagine-linked glycosylation site conforming to the consensus sequence of Asn-X-Ser, although no glycosyl residues were detected in the purified GlcNAc 2-epimerase from porcine kidney. Comparison of the nucleotide and amino acid sequences of the GlcNAc 2-epimerase with those of other known genes revealed striking similarity to the renin-binding protein (RnBP) (5, 6). The identities of the nucleotide and amino acid sequences of the enzyme to those of RnBP from porcine kidney were 99.6 and 99.0%, respectively (4). Further-

more, the amino acid sequence of the enzyme was highly homologous to those of other RnBPs from human and rat kidneys, the identities being 87.8 and 83.1%, respectively (7). That GlcNAc 2-epimerase is a RnBP was further confirmed by its ability to bind porcine kidney renin and to inhibit renin activity (4). GlcNAc 2-epimerase, as a RnBP, possesses unique hydrodynamic features such as a leucine zipper motif, which was identified as a key structure for the formation of a heterodimer of renin and one of the subunits of RnBP (8). Therefore, the crystal structure of GlcNAc 2-epimerase was required to clarify the underlying mechanisms of not only the GlcNAc 2-epimerase reaction, but also the renin-binding ability. In this paper, we report the crystallization of GlcNAc 2-epimerase from porcine kidney and some results of preliminary crystallographic characterization.

GlcNAc 2-epimerase from porcine kidney was expressed in *E. coli* and purified by the method previously described (4). The hanging drop vapor-diffusion method was used for crystallization. Protein solutions were prepared in 10 mM Tris-HCl buffer (pH 7.2). Polyethylene glycol (PEG) and ammonium acetate were used as precipitant agents. Droplets (10 μl) of a protein solution (5 mg/ml) comprising 5.5% (w/v) PEG #6000, 100 mM ammonium acetate, and 50 mM sodium citrate buffer (pH 5.0) were pipetted onto plastic coverslips, and then vapor-equilibrated at 20°C with a reservoir solution (1.0 ml) comprising 11% (w/v) PEG #6000, 200 mM ammonium acetate, and 100 mM sodium citrate buffer (pH 5.0). Prismatic, colorless crystals appeared after 20 days incubation and grew to a maximum size of 0.8 mm (Fig. 1).

A crystal was mounted in a quartz capillary with a trace amount of mother liquor. Still and precession photographs were taken at room temperature with a Nonius precession camera using Ni-filtered CuKα radiation, which was generated with a Rigaku X-ray generator operating at 40 kV and

[†] To whom correspondence should be addressed. Phone: +81-774-32-3111 (Ext.2735), Fax: +81-774-33-3004

Abbreviations: GlcNAc, *N*-acetyl-D-glucosamine; ManNAc, *N*-acetyl-D-mannosamine; PEG, polyethylene glycol.

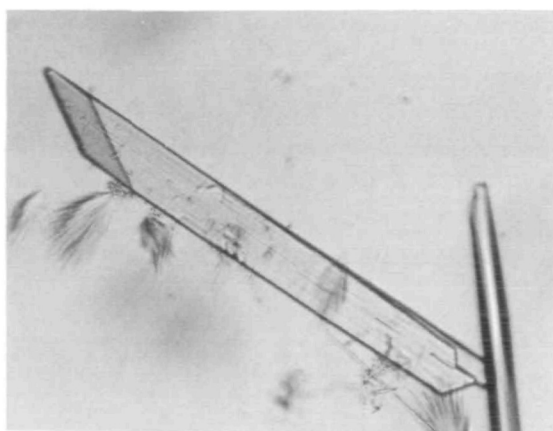


Fig. 1. A single crystal of GlcNAc 2-epimerase.

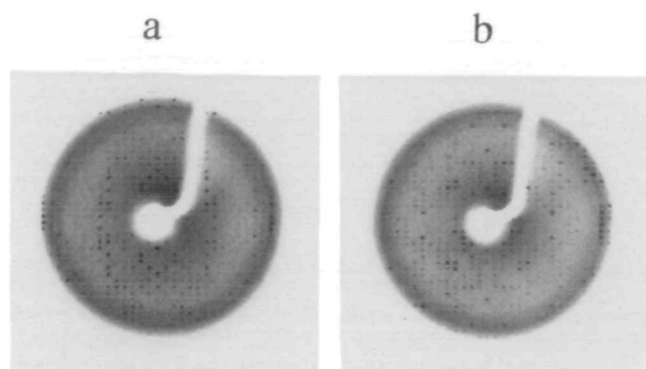


Fig. 2. Precession photographs of a crystal of GlcNAc 2-epimerase. The crystal-to-film distance was 60 mm, the precession angle 8.0 degrees and the exposure time 8 h. a, $h k 0$ zone; b, $h 0 l$ zone.

20 mA. The crystal-to-film distance was set at 60 mm. On still photographs, which were obtained by exposure for 20 min, reflections extended to at least 2.0 Å resolution and did not decay for more than 72 h. Precession photographs (Fig. 2) showed that the crystal belonged to the orthorhombic system.

The systematic absence of reflections ($h00$, $0k0$, $00l$) indicated that the space group is $P2_12_12_1$. The unit-cell dimensions are $a=78.1$, $b=97.2$, and $c=100.7$ Å, with a unit-cell volume of 764,446 Å³. Assuming that there is one dimer per asymmetric unit of the cell, the crystal volume per unit molecular mass (V_m) and the solvent volume fraction (V_{so1}) were estimated to be 2.1 Å³/Da and 0.58, respectively. These values are within the ranges of those of typical protein crystals (9).

Intensity data for the native crystals were collected with a Siemens HI-STAR detector on a MAC Science M18XHF

TABLE I. Strategy and statistics of data collected.

Detector	Siemens HI-STAR
Crystals used	1
Cell dimensions	$a=78.1$, $b=97.2$, $c=100.7$ Å
Detector distance	15 cm
Step size	0.25 degree/scan
Scan period	60 s/scan
Scans	ϕ -scan 360° ($2\theta=28^\circ$, $\chi=0^\circ$) ϕ -scan 360° ($2\theta=10^\circ$, $\chi=0^\circ$) ω -scan 50° ($2\theta=28^\circ$, $\chi=45^\circ$) ω -scan 50° ($2\theta=28^\circ$, $\chi=90^\circ$)
Collected reflections	124,509
Unique reflections	49,059
Resolution	1.92 Å
R-sym	5.6%
Completeness	82.1%

rotating anode generator. Table I shows the strategy and statistics for the data collected. The diffractions up to 1.92 Å were collected with 5.6% R-sym and 82% completeness. These data indicate the good quality of the crystals for X-ray structural analysis. A search for heavy atom derivatives is now being carried out to elucidate the high-resolution structure of GlcNAc 2-epimerase.

REFERENCES

1. Van Rinsum, J., Van Dijk, W., Hooghwinkel, G.J.M., and Werda, W. (1983) Subcellular localization and tissue distribution of sialic acid precursor-forming enzymes. *Biochem. J.* **210**, 21–28
2. Ghosh, S. and Roseman, S. (1965) The sialic acids. V. *N*-Acyl-D-glucosamine 2-epimerase. *J. Biol. Chem.* **240**, 1531–1536
3. Datta, A. (1970) Regulatory role of adenosine triphosphate on hog kidney *N*-acetyl-D-glucosamine 2-epimerase. *Biochemistry* **9**, 3363–3370
4. Maru, I., Ohta, Y., Murata, K., and Tsukada, Y. (1996) Molecular cloning and identification of *N*-acetyl-D-glucosamine 2-epimerase from porcine kidney as a renin-binding protein. *J. Biol. Chem.* **271**, 16294–16299
5. Takahashi, S., Ohsawa, T., Miura, R., and Miyake, Y. (1983) Purification and characterization of renin-binding protein (RnBP) from porcine kidney. *J. Biochem.* **93**, 1583–1594
6. Inoue, H., Fukui, K., Takahashi, S., and Miyake, Y. (1990) Molecular cloning and sequence analysis of a cDNA encoding a porcine kidney renin-binding protein. *J. Biol. Chem.* **265**, 6556–6561
7. Inoue, H., Takahashi, S., Fukui, K., and Miyake, Y. (1991) Genetic and molecular properties of human and rat renin-binding proteins with reference to the function of the leucine zipper motif. *J. Biochem.* **110**, 493–500
8. Inoue, H., Takahashi, S., Fukui, K., and Miyake, Y. (1991) Leucine zipper motif in porcine renin-binding protein (RnBP) and its relationship to the formation of an RnBP-renin heterodimer and an RnBP homodimer. *J. Biol. Chem.* **266**, 11896–11900
9. Matthews, B.W. (1968) Solvent contents of protein crystals. *J. Mol. Biol.* **33**, 491–497